Surname: Ragni

Name: Enrico

E-mail: enrico.ragni@grupposandonato.it

Scopus Author ID: 6602239679

ORCID: 0000-0003-0272-7417

Researcher ID: J-4292-2015

Education

2001-2004: PhD in Cell and Molecular Biology at University of Milano.

2001: Master's Degree in Biological Sciences (110/110 cum laude) at Università degli studi di Milano.

Awards

2007: FEMS Young Scientist Scientific Grant for the FEMS Meeting: 1st International Meeting on Cell Wall Polysaccharides of Fungi and Plants, March 10 - March 14, 2007, Anglet, France

Work experiences

2017-present: Researcher at Orthopaedics Biotechnology Lab, held by Dr. Laura de Girolamo, at Istituto Ortopedico Galeazzi, Milan, Italy

2016-2017: Post-doctoral fellow at Center for Complexity and Biosystems, held by Prof. Stefano Zapperi, at Università degli Studi di Milano, Milan, Italy

2011-2016: Research fellow at Laboratory of Regenerative Medicine - Cell Factory, held by Dr. Lorenza Lazzari, at Fondazione IRCCS Ca' Granda Ospedale Maggiore Policlinico, Milan, Italy

2006-2011: Post-doctoral fellow at Molecular Biology of Fungal Pathogens and Biotechnology Applications held by Prof. Laura Popolo, at Università degli Studi di Milano, Milan, Italy

2004-2006: Post-doctoral fellow at Cell Chemistry Lab, held by Prof. Sabine Strahl, at University of Heidelberg, Heidelberg, Germany

Lab Skills

Human cell culturing and handling, manipulation and freezing; Human Mesenchymal Stem Cells (hMSC) isolation from different tissues (bone marrow, adipose tissue, Wharton's Jelly, Amniotic Fluid, Umbilical Cord Blood, Synovial fluid); hMSC maintenance, differentiation, culturing and storage; Primary human cell lines isolation, culturing and storage; Cancer stem cells culturing; Hematopoietic Sem Cell (CD34+ and CD133+) isolation and culturing; Extracellular Vesicles isolation and characterization by nanoparticle tracking analysis, flow cytometry, western blot and qRT-PCR; DNA and RNA isolation and analysis techniques; microRNA isolation and analysis; gene manipulation; gene amplification by PCR; Molecular cloning and vector construction; qRT-PCR data generation and analysis; bioinformatics analysis and interpretation of microarray data; Flow cytometry analysis; ELISA and fluorescence techniques; Immunohistochemistry; polyacrylamide gel electrophoresis techniques and Western Blotting; advanced microscopy analysis with semiconfocal instrumentation; subcellular fractionation, isolation of the membrane fractions; Cell-Free expression of membrane proteins; expression, secretion and purification of heterologous proteins in P. pastoris and E. coli; pulse-chase labelling and immunoprecipitation techniques; HPAEC techniques; yeast genetic techniques (tetrad dissection, isolation of mutants); antifungal agent inhibition assay; evolutionary analysis of enzyme families; synthetic lethality screening in yeast by robotics instrumentation